

Determining the Effect of Galectin Insertion on Membrane Organization



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Background

The role of lectin protein and glycolipid interactions are a current and emerging interest in biochemistry due to their implications in biological functioning and pathology. Lectins are glycan-binding proteins that exist on the extracellular surface of membranes and recognize glycoconjugates on neighboring cells. Galectins more specifically exhibit conserved β -galactoside-binding sites unique to this family of lectins. Glycolipids are lipids that are covalently bound to sugar molecules at their head group and protrude from the cell membrane. Gangliosides are glycolipids covalently bound to complex oligosaccharides containing a sialic group. The use of specific saccharides at the cell surface by glycolipids to convey information is referred to as the ‘sugar code’ or glycode. This code is read by lectin proteins to induce cellular changes. Lectins have extremely stereospecific carbohydrate binding domains that allow for lectins to bind with only a very specific glycolipid. This incredible binding specificity and different combinations of glycolipids per cluster allows for many complex signals to be read and interpreted by lectins, and translated into cellular change. The cellular changes regulated by the sugar code can have pathological implications when mutations and environmental conditions alter the lectin/glycolipid interactions causing cellular dysfunction. Lectins themselves are involved in inflammation, immune responses, cell migration, autophagy, and signaling. Their mutations are linked to heart disease, cystic fibrosis, cancer, as well as many other neurological diseases.

Methods /Techniques

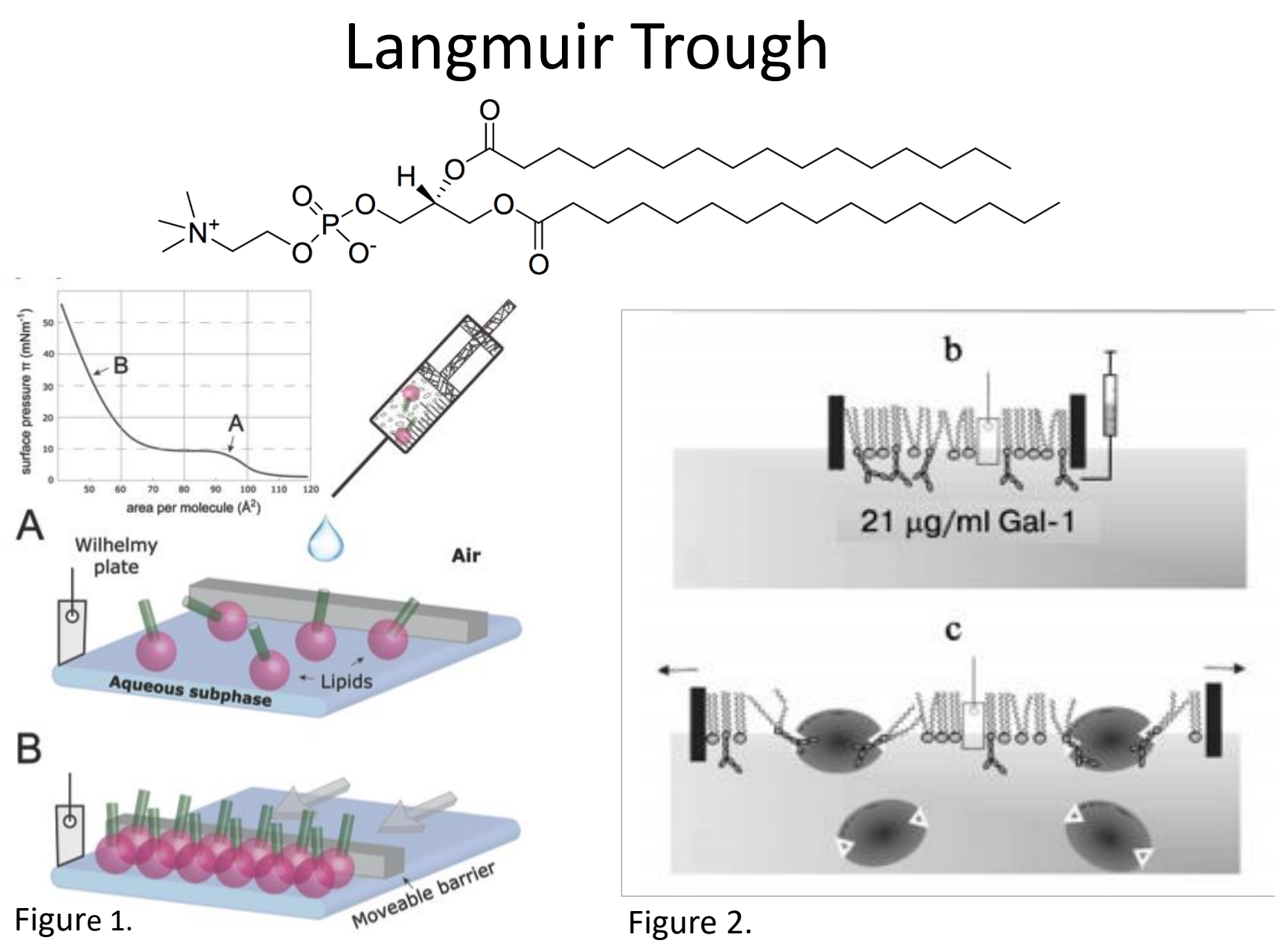
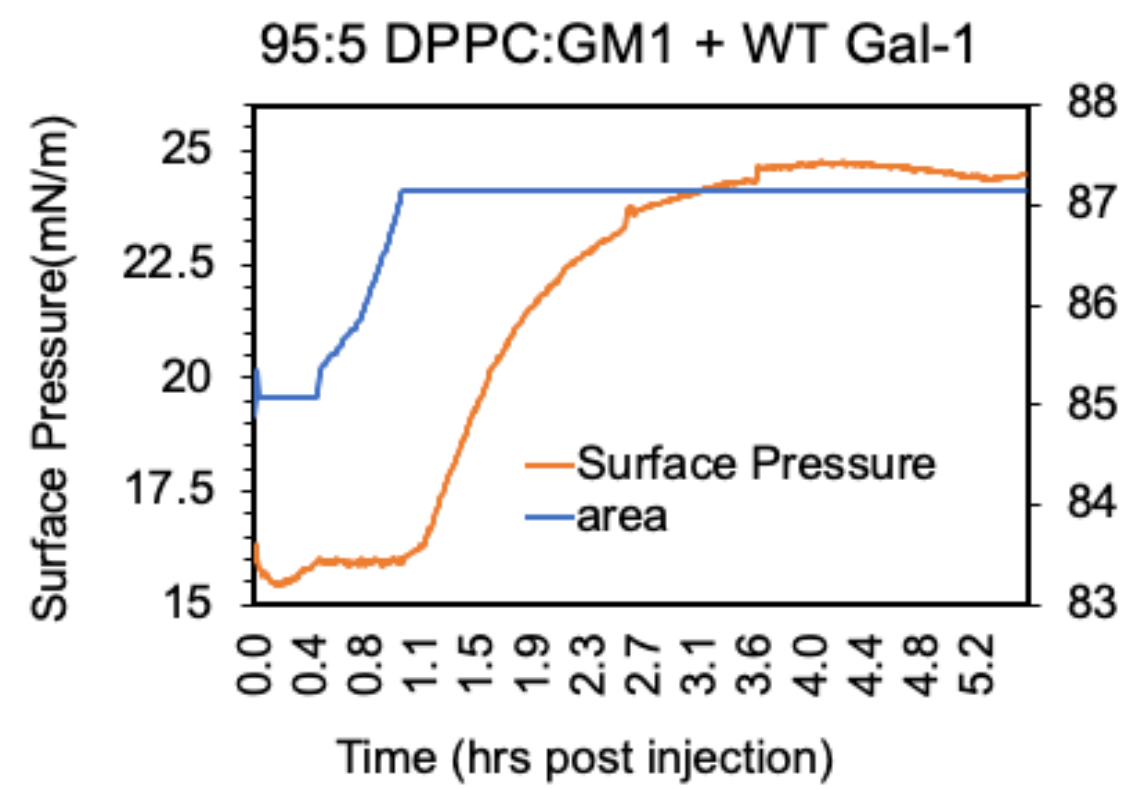


Figure 1. Gangliosides deposited onto a liquid surface, will self-organize into 2D, monolayer film, known as a Langmuir monolayer. (Figure 1 and Figure 2). DPPC lipid above.

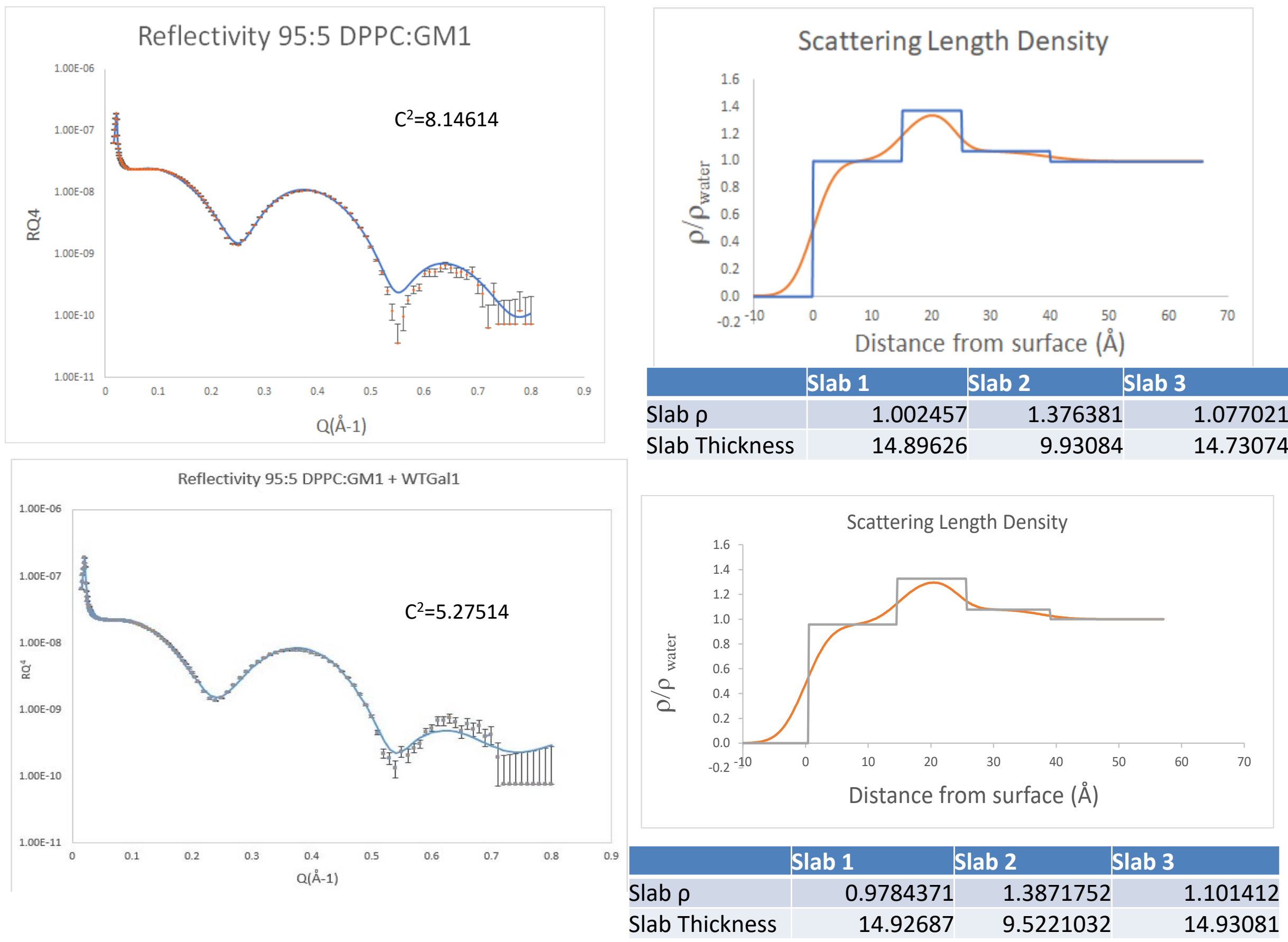
Results

Langmuir Trough – WT Gal-1 Insertion Into Membrane



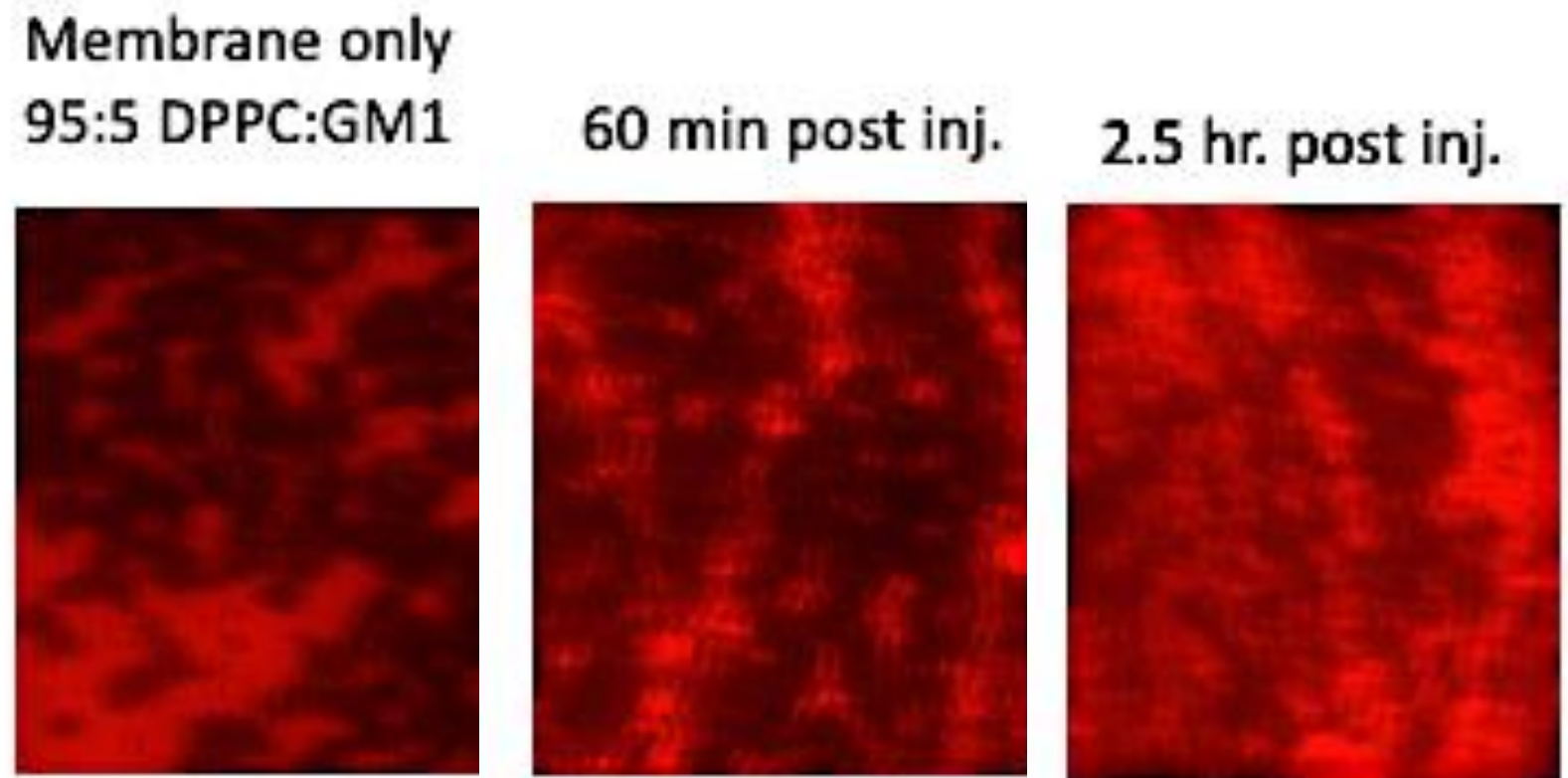
WT Gal-1 inserts into the membrane, evidenced by increase in surface pressure under constant area conditions.

X-Ray Reflectivity – WT Gal-1 Inserts Into 95:5 DPPC:GM1



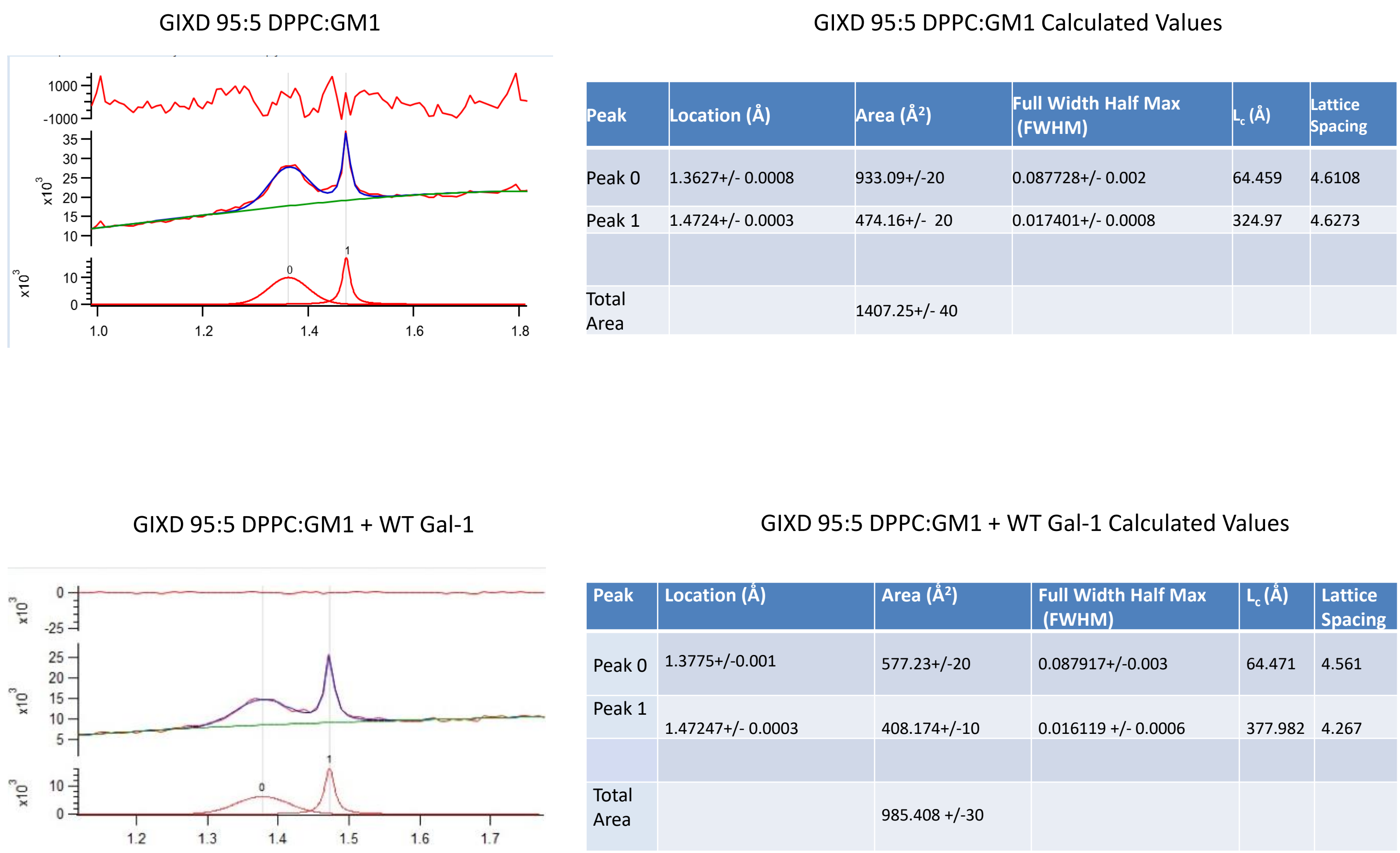
WT Gal-1 inserts into head groups of monolayer containing 95:5 DPPC:GM1.

Fluorescence Microscopy of Langmuir Film – WT Gal-1 Shrinks Condensed Domains



WT Gal-1 causes reduction in LC domain size. Liquid expanded (LE) is fluorescent red, and liquid condensed (LC) is seen as black patches.

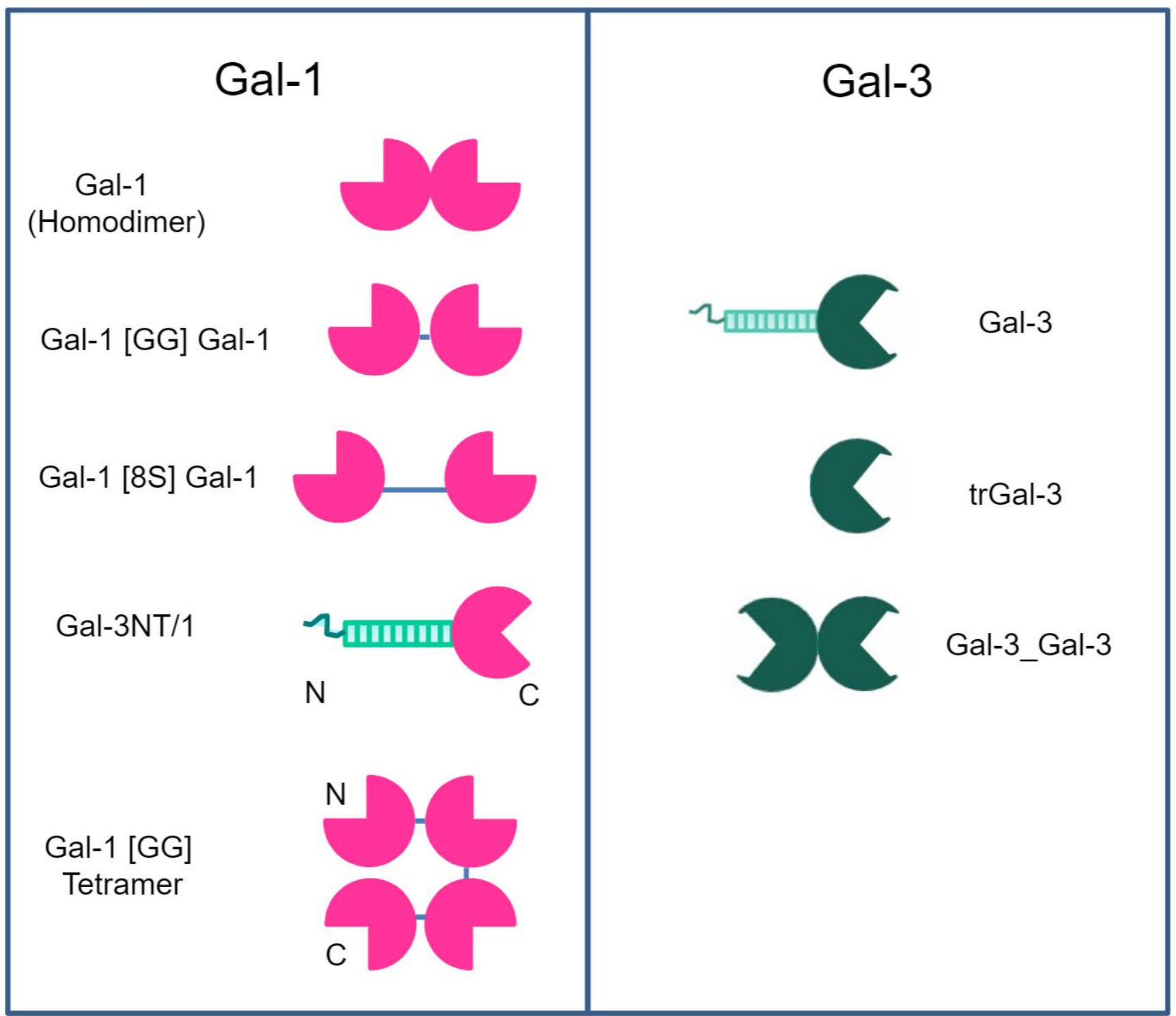
Grazing Incidence X-ray Diffraction – WT Gal-1 Impacts Lipid Packing



WT Gal-1 reorganizes lipid packing structure of 95:5 DPPC:GM1 membrane

Discussion

- Gal-1 insertion into the 95:5 DPPC:GM1 membrane yields membrane reorganization.
- Data collected from multiple experiments verifies interactions with the lipid head groups and galectin protein.
- Further analysis of different mutant galectin proteins in different membrane conditions using collected XRD data will determine differences in membrane reorganization correlating to different galectin structure.



References and Acknowledgements

References:

Majewski, J., André, S., Jones, E., Chi, E., and Gabius, H.-J. (2015) *Biochemistry (Moscow)* 80, 943–956.

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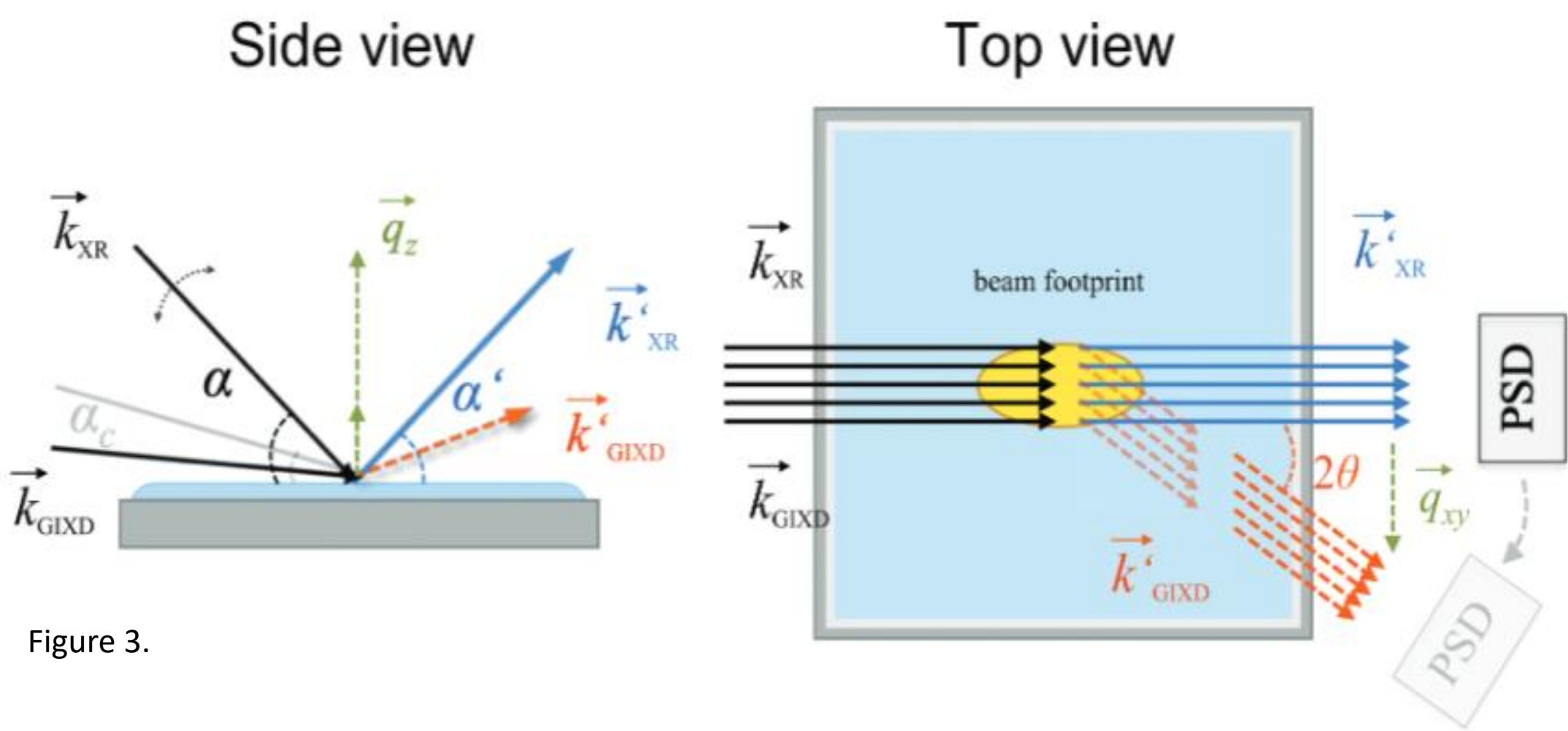
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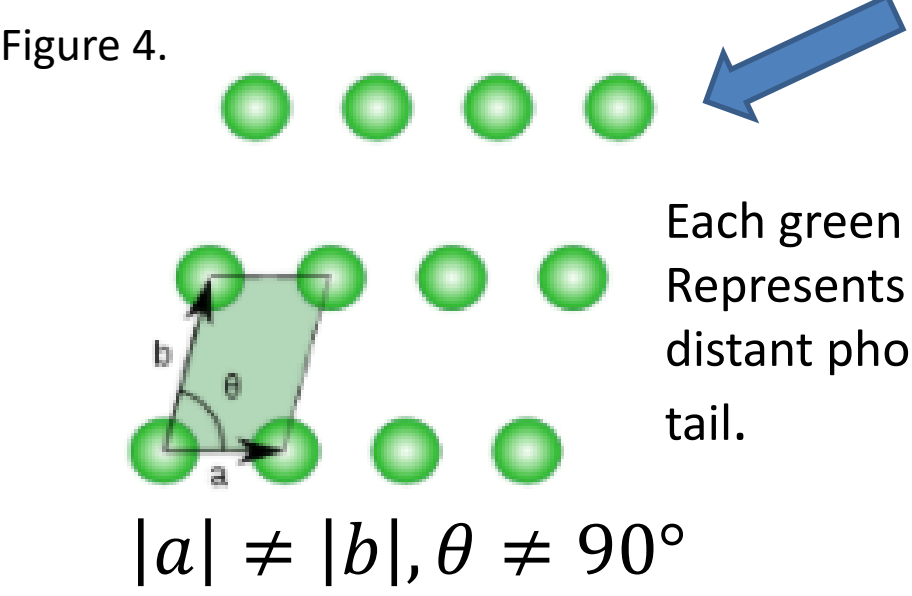
Liquid Surface X-Ray Scattering



X-ray Reflectivity (XR): measurements yield information about the out-of-plane (vertical) monolayer structure.
Grazing Incidence X-ray diffraction (GIXD): measurements provide structural in-plane information of the monolayer

Lipids Pack Into Oblique Unit Cell

Oblique (monoclinic) Unit Cell



Equations for calculations in Tables D and H

$$\text{Lattice Spacing} = \frac{2\pi}{q_{xy}}$$

Spacing in between each lipid molecule

$$L_c = \frac{0.9 \times 2\pi}{FWHM}$$

Length of coherently scattering object

In (figure 4) we see an oblique unit cell with that shows the two different lengths of $\rightarrow a$ and $\rightarrow b$, which correlate to the two different diffraction lengths and q_{xy} values used to determine the peak positions of lipids arranged in a semi crystalline manner.